

JMJ-Group – Microbiology – BMB – SDU

Title: Primer design for gene Knock Out (KO) in pathogenic *E.coli*

SOP number: SOP00XX_v01

Version number: 01

Date issued: 2013.12.03

Review date: 2014.12.03

Written by: Tina Kronborg

1. Purpose

To knock out specific genes in pathogenic *E. coli*

2. Area of application

This procedure is valid for some pathogenic *E. coli* strains (AIEC, UPEC, ETEC)

3. Apparatus and equipment

Program	Web add	Used for
ECOCYC	http://ecocyc.org/	Search for the gene of interest
Primer3	http://bioinfo.ut.ee/primer3-0.4.0/	Primer design
Bioinformatics organization Sequence Manipulation Suite (SMS) Reverse Complement	http://www.bioinformatics.org/sms/rev_comp.html or http://www.bioinformatics.org/sms2/rev_comp.html new version	Reverse and complementary the sequence

4. Materials and reagents – their shelf life and risk labelling

Name	Components (Concentrations)	Manufacturer/Cat. #	Room	Safety considerations
pKD3 fw primer (cml)	TTATACGCAAGGCGACAAGG	JMJ99 Wanner C1	V18-405a-2	20 pMol
pKD3 rev primer (cml)	GATCTTCCGTCACAGGTAGG	JMJ100 Wanner C2	V18-405a-2	20 pMol
pKD4 fw primer (kan)	CGGTGCCCTGAATGAACTGC	JMJ71 Wanner K1	V18-405a-2	20 pMol
pKD4 rev primer (kan)	CGGTGCCCTGAATGAACTGC	JMJ72 Wanner K2	V18-405a-2	20 pMol
KO Reverse primer	Made specific to the template, to KO the gene	Order at Sigma		Around 90 bp incl pKD3/4 fw primer
KO Forward primer	Made specific to the template, to KO the gene	Order at Sigma		Around 90 bp incl pKD3/4 rev primer
Primer Upstream of the gene in question	To check if the gene are KO	Order at Sigma		Around 23 bp
Primer Downstream of the gene in question	To check if the gene are KO	Order at Sigma		Around 23 bp

5. QC – Quality Control

pKD3 has Cml cassette is 1100 bp

pKD4 has Kan cassette is 1400 bp

Be aware of the size of the cassettes contra the gene in question, if they are the same size use one of the following primers together with the Up or Down primer for the gene in question

For pKD3:

JMJ99 C1 oligo, Wanner (fw)

JMJ100 C2 oligo, Wanner (rev)

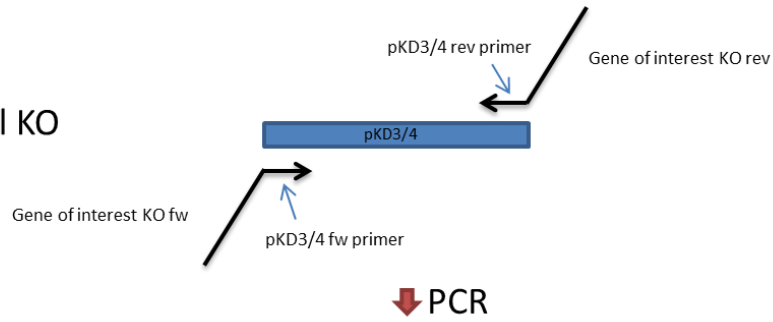
For pKD4:

JMJ71 K1 oligo, Wanner (fw)

JMJ72 K2 oligo, Wanner (rev)

If the gene I KO there will be a product with the listing primers

In common for all KO



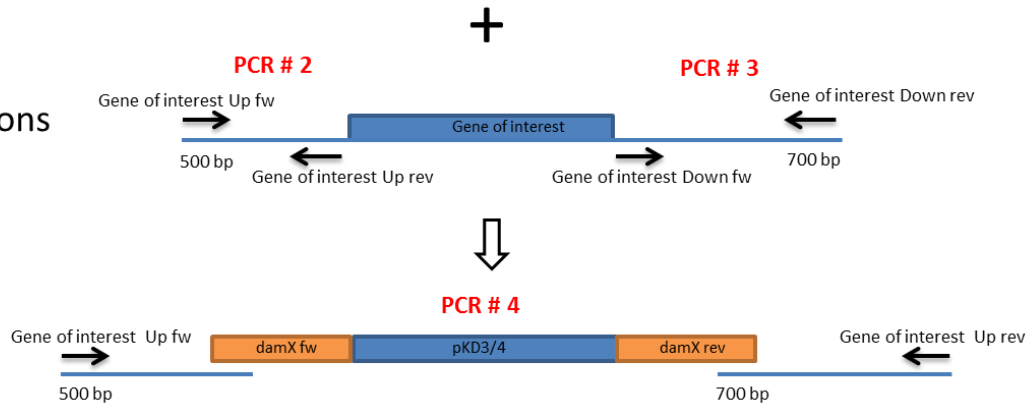
Standard KO



When KO is tricky

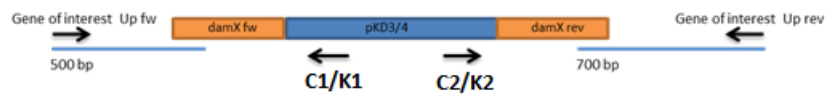


2 PCR reactions



Be aware when checking inserts!!!

C1 and K1 are reverse primers whereas C2 and K2 are forward primers!!



6. List of other SOPs relevant to this SOP

JMJ_SOP0006_v01_MM_Agarose_gel_DNA

JMJ_SOP0001_v01_MM_ON_culture_of_E.coli

JMJ_SOP0011_v01_TK_Colony_PCR_with_MyTaq

JMJ_SOP0010_v01_TK_Electroporation_of_pathogenic_E.coli_pKD46

JMJ_SOP0013_v01_TK_LA_plates_with_antibiotic

JMJ_SOP0004_v01_MM_Bacterial_freezing_stock

7. Environmental conditions required

8. Procedure

- 8.1 **Up and down primers, used to check for KO of the gene:**
- 8.2 Find the gene of interest in: <http://ecocyc.org/>
- 8.3 Change the organism to fx. UTI89
- 8.4 Use Chromosome 1
- 8.5 Type the gene name and click on the gene and chose Nucleotide Sequence, Advanced
- 8.6 Type 400 bp upstream and 400 bp downstream and show sequence
- 8.7 Copy the sequence to <http://bioinfo.ut.ee/primer3-0.4.0/>
- 8.8 Chose primer size at 23, Tm at 62 °C and the Product Size Ranges according to gene size, be sure that the primers start outside the gene.
- 8.9 The boxes with Pick left primer, or use left primer below: and Pick right primer, or use right primer below (5' to 3' on opposite strand) shall have a mark.
- 8.10 Push Pick Primers.
- 8.11 **Primers for pKD3 and pKD4 used to KO the gene with:**
- 8.12 pKD3/4 fw primer : GTGTAGGCTGGAGCTGCTTC
- 8.13 pKD3/4 rev primer: CATATGAATATCCTCCTTA
- 8.14 Find the gene of interest again.
- 8.15 Type 100 bp upstream and 100 bp downstream and show sequence
- 8.16 From the start (fw primer) and stop (rev primer) codon you pick 6 codons and then go backwards until 70 bp in all
- 8.17 Use: http://www.bioinformatics.org/sms/rev_comp.html to make the rev primer sequence reverse and complementary
- 8.18 Pair the sequences with pKD3/4 fw and rev primer respectively
- 8.19 **For difficult genes:**
- 8.20 1st set of primers – Up primers:
- 8.21 Follow 8.1 – 8.5
- 8.22 Type 500 bp upstream and show sequence
- 8.23 Copy the sequence from the 500 bp extra and into the start codon and paste it into Primer 3. It is important that the region include the sequence from the gene of interest fw primer
- 8.24 Chose a product size range from 470-480, primer size on 22 and pick primers
- 8.25 2nd set of primers – Down primers:
- 8.26 Follow 8.1 – 8.5
- 8.27 Type 700 bp downstream and show sequence
- 8.28 Copy the sequence from the 700 bp extra and into the stop codon and paste it into Primer 3. It is important that the region include the sequence from the gene of interest rev primer
- 8.29 Chose a product size range from 670-680, primer size on 22 and pick primers

9. Waste handling

Not required

Chemical name	Concentration	Type of waste (C, Z...)	Remarks

10. Time consumption

- Total-time 2 hours
- Hands-on-time 1 hour

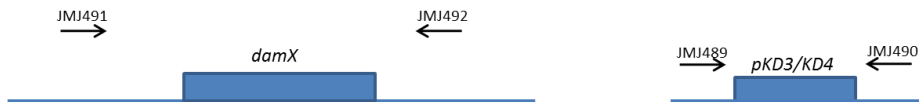
11. Scheme of development

Date / Initials	Version No.	Description of changes
14.01.15 / TK	01	The SOP has been written

12. Appendixes

Procedure with an example:

Primer design for KO



Knock out of *damX*

damX 1287 bp

JMJ489 KO_{fw}

JMJ490 KO_{rev}

JMJ491 up

JMJ492 down

PCR product sizes: 2174

JMJ489 + JMJ490 = 1100/1400bp depending on pKD3/pKD4 template

Find the gene of interest in:

<http://ecocyc.org/>

Change the organism to fx. UTI89

Use Chromosome 1

Type the gene name and click on the gene and chose Nucleotide Sequence, Advanced

Type 400 bp upstream and 400 bp downstream and show sequence

Copy the sequence to <http://bioinfo.ut.ee/primer3-0.4.0/>

Chose primer size at 23, T_m at 62 °C and the Product Size Ranges according to gene size, be sure that the primers start outside the gene.

The boxes with Pick left primer, or use left primer below: and Pick right primer, or use right primer below (5' to 3' on opposite strand) shall have a mark.

Push Pick Primers.

Ex. damX:

Nucleotide sequence:

>gnl|ECOL364106|GHPQ-3849 damX "GHPQ-3849-MONOMER"
(complement(3769974..3768688)) Escherichia coli UTI89 Chromosome 1

```

atgGATGAAT TCAAACCAGA AGACGAGCTG AAACCCGATC CCAGCGATCG TCGTACTGGT
CGTTCTCGTC AATCTTCTGA ACGTTCTGAG CGTACTGAAC GTGGCGAACC GCAGATCAAT
TTTGATGATA TTGAACTTGA TGACACTGAC GATCGCCGTC CGACTCGTGC GCAAAAAGCG
CGTAATGAGG AACCGGAAAT CGAAGAAGAA ATTGACGAAT CCGAAGATGA AACCGTGGAT
GAAGAGCGCG TAGAGCGTCC TCCGCGTAAG CGCAAAAAG CAGCCAGTAA ACCCGCTTCT
CGTCAGTATA TGATGATGGG TGTCGGCATT CTGGTTCTAC TGCTGTTGAT CATCGGTATC
GGTTCTGCGC TAAAAGCCCC CTCGACCTCT TCCAGCGATC AAACCGCGTC TGGCGAGAAG
AGTATTGATC TTGCAGGCAA TGCAGCCGAT CAGGCCAATG GCGTGCAGCC AGCGCCGGGA
ACCACGTCTG CGGAAAATAC TCAGCAGGAT GTTTCTCTGC CGCCGATCTC TTCTACGCCG
ACTCAAGGGC AAACCCCGGT GGCAACGGAT GGTCAACAAC GTGTTGAAGT GCAGGGTGAC
CTGAACAATG CGCTGACCCA GCCACAAAAT CAGCAACAGT TGAACAATGT GGCGGTCAAT
TCTACATTGC CACTGAACC CGCAACGGTT GCGCCTGTTT GCAATGGCAA TGCATCGCGT
GACACGGCGA AAACGCAAAC CGCTGAACGT CCGGCCACTA CGCGCCCGGC TCGTCAGCAG
GCGGTGATTG AACCGAAAAA ACCGCAAGCA ACCGTGAAAA GCGAACCAGAA GCCAGTCGCA
CAGACGCCGA AGCGAACTGA ACCAGCTGCC CCTGTGGCGA GCACAAAGGC ACCGGCTGCG
ACTTCTACGC CAGCACCAA AGAGACGGCG ACTACGGCTC CAGTACAGAC GGCATCCCCG
GCGCAAACCA CGGCAACACC AGCCGCTGGA GGGAAAGACCG CAGGTAATGT TGGTTCGTTG
AAATCGGCAC CGTCCAGCCA TTACTACTCTG CAGCTGAGCA GTTCCTCTAA CTACGACAAC
CTGAACGGTT GGGCGAAGAA AGAGAATCTG AAAAACTACG TTGTCTATGA AACGACGCGT
AATGGTCAGC CGTGGTATGT CTTGGTTTCT GCGCTGTATG GCGCTGTATG AGAGGCGAAA
AAAGCGGTAT CTACATTGCC AGCAGATGTC CAGGCCAAAA ACCCGTGGGC GAAACCGCTG
CGTCAGGTAC AGGCCGATCT GAAGtaa
    
```

With 400 extra bp on each side of the gene:

```

AATTGGTTAC ATGGTGAAGC GGTGCGTGGC GGTATGGTGA TGGCGGCGCG GACGTCGGAA
CGTCTCGGGC AGTTTAGTTC TGCCGAAACG CAGCGTATTA TAACCCTGCT CACGCGGGCT
GGGTACCAGG TCAATGGGCC GCGCGAAATG TCCGCGCAGG CGTATTTACC GCATATGCTG
CGTGACAAGA AAGTCCTTGC GGGAGAGATG CGCTTAATTC TTCCGTTGGC AATTGGTAAG
AGTGAAGTTC GCAGCGGCGT TTCGCACGAG CTTGTTCTTA ACGCCATTGC CGATTGTCAA
TCAGCGTAAC AACCAAGAAAG GTCAGGCCGC TTATCAAGCG GTCTATTAGC TTCAGGTTAA
TTGCAACGTG GTAAGCATTG ACCTTTTAGT GGGGTGTTAA atgGATGAAT TCAAACCAGA
AGACGAGCTG AAACCCGATC CCAGCGATCG TCGTACTGGT CGTTCTCGTC AATCTTCTGA
ACGTTCTGAG CGTACTGAAC GTGGCGAACC GCAGATCAAT TTTGATGATA TTGAACTTGA
TGACACTGAC GATCGCCGTC CGACTCGTGC GCAAAAAGCG CGTAATGAGG AACCGGAAAT
CGAAGAAGAA ATTGACGAAT CCGAAGATGA AACCGTGGAT GAAGAGCGCG TAGAGCGTCG
TCCGCGTAAG CGCAAAAAG CAGCCAGTAA ACCCGCTTCT CGTCAGTATA TGATGATGGG
TGTCGGCATT CTGGTTCTAC TGCTGTTGAT CATCGGTATC GGTTCTGCGC TAAAAGCCCC
CTCGACCTCT TCCAGCGATC AAACCGCGTC TGGCGAGAAG AGTATTGATC TTGCAGGCAA
TGGCACCAGT CAGGCCAATG GCGTGCAGCC AGCGCCGGGA ACCACGTCTG CGGAAAATAC
TCAGCAGGAT GTTTCTCTGC CGCCGATCTC TTCTACGCCG ACTCAAGGGC AAACCCCGGT
GGCAACGGAT GGTCAACAAC GTGTTGAAGT GCAGGGTGAC CTGAACAATG CGCTGACCCA
GCCACAAAAT CAGCAACAGT TGAACAATGT GGCGGTCAAT TCTACATTGC CACTGAACC
CGCAACGGTT GCGCCTGTTT GCAATGGCAA TGCATCGCGT GACACGGCGA AAACGCAAAC
CGCTGAACGT CCGGCCACTA CGCGCCCGGC TCGTCAGCAG GCGGTGATTG AACCGAAAAA
ACCGCAAGCA ACCGTGAAAA CGGAACCAGAA GCCAGTCGCA CAGACGCCGA AGCGAACTGA
ACCAGCTGCC CCTGTGGCGA GCACAAAGGC AACCGGCTGCG ACTTCTACGC CAGCACCAA
AGAGACGGCG ACTACGGCTC CAGTACAGAC GGCATCCCCG GCGCAAACCA CGGCAACACC
AGCCGCTGGA GGGAAAGACCG CAGGTAATGT TGGTTCGTTG AAATCGGCAC CGTCCAGCCA
TTACTACTCT CAGCTGAGCA GTTCCTCTAA CTACGACAAC CTGAACGGTT GGGCGAAGAA
AGAGAATCTG AAAAACTACG TTGTCTATGA AACGACGCGT AATGGTCAGC CGTGGTATGT
CCTGGTTTCT GCGCTGTATG CTTCGAAAAG AGAGGCGAAA AAAGCGGTAT CTACATTGCC
AGCAGATGTC CAGGCCAAAA ACCCGTGGGC GAAACCGCTG CGTCAGGTAC AGGCCGATCT
    
```


601 CGAAGAAGAAATTGACGAATCCGAAGATGAAACCGTGGATGAAGAGCGCGTAGAGCGTCG
661 TCCGCGTAAGCGCAAAAAAGCAGCCAGTAAACCCGCTTCTCGTCAGTATATGATGATGGG
721 TGTCGGCATTCTGGTTCTACTGCTGTTGATCATCGGTATCGGTTCTGCGCTAAAAGCCCC
781 CTCGACCTCTTCCAGCGATCAAACCGCGTCTGGCGAGAAGAGTATTGATCTTGCAGGCAA
841 TGCGACCGATCAGGCCAATGGCGTGCAGCCAGCGCCGGAACCACGTCTGCGGAAAATAC
901 TCAGCAGGATGTTTCTCTGCCGCCGATCTCTTCTACGCCGACTCAAGGGCAAACCCCGGT
961 GGCAACGGATGGTCAACAACGTGTTGAAGTGCAGGGTGACCTGAACAATGCGCTGACCCA
1021 GCCACAAAATCAGCAACAGTTGAACAATGTGGCGGTCAATTCTACATTGCCGACTGAACC
1081 CGCAACGGTTGCGCCTGTTTCGCAATGGCAATGCATCGCGTGACACGGCGAAAACGCAAAC
1141 CGCTGAACGTCCGGCCACTACGCGCCCGGCTCGTCAGCAGGCGGTGATTGAACCGAAAAA
1201 ACCGCAAGCAACCGTGAAAACGGAACCGAAGCCAGTCGCACAGACGCCGAAGCGAACTGA
1261 ACCAGCTGCCCCGTGGCGAGCACAAAGGCACCGGCTGCGACTTCTACGCCAGCACCAAA
1321 AGAGACGGCGACTACGGCTCCAGTACAGACGGCATCCCCGGCGCAAACCACGGCAACACC
1381 AGCCGCTGGAGGGAAGACCGCAGGTAATGTTGGTTTCGTTGAAATCGGCACCGTCCAGCCA
1441 TTACTACTCTGCAGCTGAGCAGTTCCTCTAACTACGACAACCTGAACGGTTGGGCGAAGAA
1501 AGAGAATCTGAAAACTACGTTGTCTATGAAACGACGCGTAATGGTCAGCCGTGGTATGT
1561 CCTGGTTTCTGGCGTGTATGCTTCGAAAGAAGAGGCGAAAAAAGCGGTATCTACATTGCC
1621 AGCAGATGTCCAGGCCAAAACCCGTGGGCGAAACCGCTGCGTCAGGTACAGGCCGATCT
1681 GAAG_taaTCAAGGTTATCTCCCGCAATGGTTTATCATTGCGGGCGTTGCCTGAAGCGCTG
1741 GATGCTGTCCGAGCTTCTCCACAGCCGGAGAAGGTGTAATTAGTTAGTCAGCATGAAGA


```

Left      929      0      0      0      3      0      102      693      0      1      0      30
100
Right     916      0      0      0      0      0      257      398      0      2      0      33
222

```

Pair Stats:

considered 50, unacceptable product size 41, high end compl 1, ok 8
 primer3 release 1.1.4

So **damX up** is:

AACGTCTCGGGCAGTTTAGTTCT

And **damX down** is:

TCGCGGAACTGATAGTAAACCTC

It will give a product size on 2174 bp

Primers for pKD3 and pKD4 and damX:

pKD3/4 fw primer : **GTGTAGGCTGGAGCTGCTTC**

pKD3/4 rev primer: **CATATGAATATCCTCCTTA**

Find the gene of interest again.

Type 100 bp upstream and 100 bp downstream and show sequence

Organism: *Escherichia coli* UTI89

Chromosome: **Chromosome 1**

Region: 3768588 - 3770074 (reverse complement)

```

TCAGCGTAAC  AACAAGAAAG  GTCAGGCCGC  TTATCAAGCG  GTCTATTAGC  TTCAGGTTAA
TTGCAACGTG  GTAAGCATTA  ACCTTTTAGT  GGGGTGTTAA  atgGATGAAT  TCAAACCAGA
AGACGAGCTG  AAACCCGATC  CCAGCGATCG  TCGTACTGGT  CGTTCTCGTC  AATCTTCTGA
ACGTTCTGAG  CGTACTGAAC  GTGGCGAACC  GCAGATCAAT  TTTGATGATA  TTGAACTTGA
TGACACTGAC  GATCGCCGTC  CGACTCGTGC  GCAAAAAGCG  CGTAATGAGG  AACCGGAAAT
CGAAGAAGAA  ATTGACGAAT  CCGAAGATGA  AACCGTGGAT  GAAGAGCGCG  TAGAGCGTCG
TCCGCGTAAG  CGCAAAAAG  CAGCCAGTAA  ACCCGCTTCT  CGTCAGTATA  TGATGATGGG
TGTCGGCATT  CTGGTTCTAC  TGCTGTTGAT  CATCGGTATC  GGTTCGCGC  TAAAAGCCCC
CTCGACCTCT  TCCAGCGATC  AAACCGCGTC  TGGCGAGAAG  AGTATTGATC  TTGCAGGCAA
TGCGACCGAT  CAGGCCAATG  GCGTGCAGCG  AGCGCCGGGA  ACCACGTCTG  CGGAAAATAC
TCAGCAGGAT  GTTTCTCTGC  CGCCGATCTC  TTCTACGCCG  ACTCAAGGGC  AAACCCCGGT
GCACAACGGAT  GGTCAACAAC  GTGTTGAAGT  GCAGGGTGAC  CTGAACAATG  CGCTGACCCA
GCCACAAAAT  CAGCAACAGT  TGAACAATGT  GGCGGTCAAT  TCTACATTGC  CGACTGAACC
CGCAACGGTT  GCGCCTGTTT  GCAATGGCAA  TGCATCGCGT  GACACGGCGA  AAACGCAAAC
CGCTGAACGT  CCGGCCACTA  CGCGCCCGGC  TCGTCAGCAG  GCGGTGATTG  AACCGAAAAA
ACCGCAAGCA  ACCGTGAAAA  CGGAACCGAA  GCCAGTCGCA  CAGACGCCGA  AGCGAACTGA
ACCAGCTGCC  CCTGTGGCGA  GCACAAAGGC  ACCGGCTGCG  ACTTCTACGC  CAGCACCAAA
AGAGACGGCG  ACTACGGCTC  CAGTACAGAC  GGCATCCCCG  GCGCAAACCA  CGGCAACACC

```

```
AGCCGCTGGA GGG AAGACCG CAGGTAATGT TGGTTCGTTG AAATCGGCAC CGTCCAGCCA
TTACTACTCTG CAGCTGAGCA GTTCCTCTAA CTACGACAAC CTGAACGGTT GGGCGAAGAA
AGAGAATCTG AAAAACTACG TTGTCTATGA AACGACGCGT AATGGTCAGC CGTGGTATGT
CCTGGTTTCT GCGGTGTATG CTTTCGAAAGA AGAGGCGAAA AAAGCGGTAT CTACATTGCC
AGCAGATGTC CAGGCCAAAA ACCCGTGGGC GAAACCGCTG CGTCAGGTAC AGGCCGATCT
GAAGtaaTCA AGGTTATCTC CCGCAATGGT TTATCATTGC GGGCGTTGCC TGAAGCGCTG
GATGCTGTCG GAGCTTTCTC CACAGCCGGA GAAGGTGTAA TTAGTTA
```

From the start and stop codon you pick 6 codons and then go backwards until 70 bp in all

For damX fw it will give:

```
GCTTCAGGTTAATTGCAACGTGGTAAGCATTAACTTTTAGTGGGGTGTTAAatgGATGAAT
TCAAACCA
```

And for damX rev:

```
CAGGCCGATCTGAAGtaaTCAAGGTTATCTCCCGCAATGGTTTATCATTGCGGGCGTTGCCT
GAAGCGCT
```

Use:

http://www.bioinformatics.org/sms/rev_comp.html

to make the sequence reverse and complementary:

```
AGCGCTTCAGGCAACGCCCGCAATGATAAACCATTGCGGGAGATAACCTTGAttaCTTCAGA
TCGGCCTG
```

So damX KO fw is:

```
GCTTCAGGTTAATTGCAACGTGGTAAGCATTAACTTTTAGTGGGGTGTTAAatgGATGAAT
TCAAACCA GTGTAGGCTGGAGCTGCTTC
```

So damX KO rev is:

```
AGCGCTTCAGGCAACGCCCGCAATGATAAACCATTGCGGGAGATAACCTTGAttaCTTCAGA
TCGGCCTG CATATGAATATCCTCCTTA
```

If it is difficult to knock out the gene it can be helpful to make the PCR product around 1000 bp longer in all. You can do so by making 2 more set of primers and run additional 2 PCR

1th set of primers:

Find the gene in question, in Nucleotide Sequence, Advanced type 500 bp upstream and show sequence. Copy the sequence from the 500 bp extra and into the start codon (marked with yellow) and paste it into Primer 3. It is important that the region include the sequence from the damX fw primer.

```

AGAAGTTGTC GCGGCCGACG AGCGCGAAAC CGGGTTACGT GCTTTACTGA ATCTGGGACA
CACCTTTGGT CATGCCATTG AAGCTGAAAT GGGGTATGGC AATTGGTTAC ATGGTGAAGC
GGTCGCTGCG GGTATGGTGA TGGCGGCGCG GACGTCGGAA CGTCTCGGGC AGTTTAGTTC
TGCCGAAACG CAGCGTATTA TAACCCTGCT CACGCGGGCT GGGTTACCGG TCAATGGGCC
GCGCGAAATG TCCGCGCAGG CGTATTTACC GCATATGCTG CGTGACAAGA AAGTCCTTGC
GGGAGAGATG CGCTTAATTC TTCCGTTGGC AATTGGTAAG AGTGAAGTTC GCAGCGGCGT
TTCGCACGAG CTTGTTCTTA ACGCCATTGC CGATTGTCAA TCAGCGTAAC AACAAAGAAAG
GTCAGGCCGC TTATCAAGCG GTCTATTAGC TTCAGGTTAA TTGCAACGTG GTAAGCATTAA
ACCTTTTAGT GGGGTGTTAA atgGATGAAT TCAAACCAGA AGACGAGCTG AAACCCGATC
CCAGCGATCG TCGTACTGGT CGTTCTCGTC AATCTTCTGA ACGTTCTGAG CGTACTGAAC
GTGGCGAACC GCAGATCAAT TTTGATGATA TTGAACTTGA TGACACTGAC GATCGCCGTC
CGACTCGTGC GCAAAAAGCG CGTAATGAGG AACC GGAAAT CGAAGAAGAA ATTGACGAAT
CCGAAGATGA AACCCTGGAT GAAGAGCGCG TAGAGCGTCC TCCGCGTAAG CGCAAAAAAG
CAGCCAGTAA ACCCGTCTCT CGTCAGTATA TGATGATGGG TGTCGGCATT CTGGTCTAC
TGCTGTTGAT CATCGGTATC GGTTCCTGCGC TAAAAGCCCC CTCGACCTCT TCCAGCGATC
AAACCGCGTC TGGCGAGAAG AGTATTGATC TTGCAGGCAA TCGGACCGAT CAGGCCAATG
GCGTGCAGCC AGCGCCGGGA ACCACGTCTG CGGAAAATAC TCAGCAGGAT GTTCTCTGC
CGCCGATCTC TTCTACGCCG ACTCAAGGGC AAACCCCGGT GGCAACGGAT GGTCACAAC
GTGTTGAAGT GCAGGGTGAC CTGAACAATG CGCTGACCCA GCCACAAAAT CAGCAACAGT
TGAACAATGT GGCGGTCAAT TCTACATTGC CGACTGAACC CGCAACGGTT GCGCCTGTTT
GCAATGGCAA TGCATCGCGT GACACGGCGA AAACGCAAAC CGCTGAACGT CCGGCCACTA
CGCGCCCGGC TCGTCAGCAG GCGGTGATTG AACC GAAAAA ACCGCAAGCA ACCGTGAAAA
CGGAACCGAA GCCAGTCGCA CAGACGCCGA AGCGAACTGA ACCAGCTGCC CCTGTGGCGA
GCACAAAGGC ACCGGCTGCG ACTTCTACGC CAGCACAAA AGAGACGGCG ACTACGGCTC
CAGTACAGAC GGCATCCCCG GCGCAAACCA CGGCAACACC AGCCGCTGGA GGGAAAGACCG
CAGGTAATGT TGGTTCGTTG AAATCGGCAC CGTCCAGCCA TTACACTCTG CAGCTGAGCA
GTTCCCTCTAA CTACGACAAC CTGAACGGTT GGGCGAAGAA AGAGAATCTG AAAA ACTACG
TTGTCTATGA AACGACCGGT AATGGTCAGC CGTGGTATGT CCTGGTTTCT GGCCTGTATG
CTTCGAAAGA AGAGGCGAAA AAAGCGGTAT CTACATTGCC AGCAGATGTC CAGGCCAAAA
ACCCGTGGGC GAAACCGCTG CGTCAGGTAC AGGCCGATCT GAAGtaa
    
```

Chose a product size range from 470-480, primer size on 22 and pick primers

No mispriming library specified
Using 1-based sequence positions

OLIGO	start	len	tm	gc%	any	3' seq
LEFT PRIMER	31	22	60.41	45.45	4.00	2.00 CGGGTTACGTGCTTTACTGAAT
RIGHT PRIMER	509	22	60.46	40.91	4.00	1.00 TTCATCcatTTAACACCCCACT

SEQUENCE SIZE: 510

RIGHT PRIMER 496 22 59.77 45.45 4.00 2.00
 CACCCCACTAAAAGGTTAATGC
 PRODUCT SIZE: 470, PAIR ANY COMPL: 4.00, PAIR 3' COMPL: 2.00

Statistics

	con sid ered	too many Ns	in tar get	in excl reg	bad GC%	no GC clamp	tm too low	tm too high	high any compl	high 3' compl	poly X	high end stab
ok												
Left	383	0	0	0	4	0	42	288	0	2	0	4
Right	402	0	0	0	0	0	147	83	0	0	0	3

Pair Stats:
 considered 366, unacceptable product size 355, high end compl 2, ok 9
 primer3 release 1.1.4

damX up fw: CGGGTTACGTGCTTTACTGAAT
damX up rev: TTCATCcatTTAACACCCCACT

2nd set of primers

Find the gene in question, in Nucleotide Sequence, Advanced type 700 bp downstream and show sequence. Copy the sequence from the 700 bp extra and into the stop codon (marked with yellow) and paste it into Primer 3. It is important that the region include the sequence from the damX rev primer.

```

atgGATGAAT TCAAACCAGA AGACGAGCTG AAACCCGATC CCAGCGATCG TCGTACTGGT
CGTTCCTCGTC AATCTTCTGA ACGTTCTGAG CGTACTGAAC GTGGCGAACC GCAGATCAAT
TTTGATGATA TTGAACTTGA TGACACTGAC GATCGCCGTC CCGACTCGTGC GCAAAAAGCG
CGTAATGAGG AACCGGAAAT CGAAGAAGAA ATTGACGAAT CCGAAGATGA AACCGTGGAT
GAAGAGCGCG TAGAGCGTCG TCCGCGTAAG CGCAAAAAG CAGCCAGTAA ACCCGTTCT
CGTCAGTATA TGATGATGGG TGTCGGCATT CTGGTTCTAC TGCTGTTGAT CATCGGTATC
GGTTCCTGCGC TAAAAGCCCC CTCGACCTCT TCCAGCGATC AAACCCGCGTC TGGCGAGAAG
AGTATTGATC TTGCAGGCAA TGCGACCGAT CAGGCCAATG GCGTGCAGCC AGCGCCGGGA
ACCACGTCTG CGGAAAATAC TCAGCAGGAT GTTTCTCTGC CGCCGATCTC TTCTACGCCG
ACTCAAGGGC AAACCCCGGT GGCAACGGAT GGTCAACAAC GTGTTGAAGT GCAGGGTGAC
CTGAACAATG CGCTGACCCA GCCACAAAAT CAGCAACAGT TGAACAATGT GGCGGTCAAT
TCTACATTGC CGACTGAACC CGCAACGGTT GCGCCTGTTT GCAATGGCAA TGCATCGCGT
GACACGGCGA AAACGCAAAC CGCTGAACGT CCGGCCACTA CGCGCCCGGC TCGTCAGCAG
GCGGTGATTG AACCGAAAAA ACCGCAAGCA ACCGTGAAAA CGGAACCGAA GCCAGTCGCA
CAGACGCCGA AGCGAACTGA ACCAGCTGCC CCTGTGGCGA GCACAAAGGC ACCGGTGGC
ACTTCTACGC CAGCACAAA AGAGACGGC ACTACGGCTC CAGTACAGAC GGCATCCCCG
GCGCAAACCA CGGCAACACC AGCCGCTGGA GGGAAAGACCG CAGGTAATGT TGGTTCGTTG
AAATCGGCAC CGTCCAGCCA TTACTACTCTG CAGCTGAGCA GTTCCTCTAA CTACGACAAC
CTGAACGGTT GGGCGAAGAA AGAGAATCTG AAAAACTACG TTGTCTATGA AACGACGCGT
AATGGTCAGC CGTGGTATGT CCTGGTTTTCT GGCGTGTATG CTTCGAAAGA AGAGGCGAAA
AAAGCGGTAT CTACATTGCC AGCAGATGTC CAGGCCAAAA ACCCGTGGGC GAAACCGCTG
CGTCAGGTAC AGCCGATCT GAAGTaaTCA AGGTTATCTC CCGCAATGGT TTATCATTGC
GGGCGTTGCC TGAAGCGCTG GATGCTGTCG GAGCTTTCTC CACAGCCGGA GAAGGTGTAA
TTAGTTAGTC AGCATGAAGA AAAATCGCGC TTTTTTGAAG TGGGCAGGGG GCAAGTATCC
CCTGCTTGAT GATATTAAC GGCATTTGCC CAAGGGCGAA TGTCTGGTTG AGCCTTTTGT
AGGTGCCGGG TCGGTGTTTT TCAACACCGA CTTTTCTCGT TATATCCTTG CCGATATCAA
  
```


KEYS (in order of precedence):

>>>>> left primer
 <<<<<< right primer

ADDITIONAL OLIGOS

	<u>start</u>	<u>len</u>	<u>tm</u>	<u>gc%</u>	<u>any</u>	<u>3'</u>	<u>seq</u>
1 LEFT PRIMER	14	22	60.17	40.91	3.00	1.00	
TTATCTCCCGCAATGGTTTATC							
RIGHT PRIMER	691	19	60.61	52.63	4.00	2.00	AAGTTGGCAGTCGCAGACA
PRODUCT SIZE: 678, PAIR ANY COMPL: 3.00, PAIR 3' COMPL: 2.00							
2 LEFT PRIMER	14	22	60.17	40.91	3.00	1.00	
TTATCTCCCGCAATGGTTTATC							
RIGHT PRIMER	692	20	61.97	50.00	4.00	2.00	AAAGTTGGCAGTCGCAGACA
PRODUCT SIZE: 679, PAIR ANY COMPL: 3.00, PAIR 3' COMPL: 2.00							
3 LEFT PRIMER	15	22	61.51	40.91	3.00	2.00	
TATCTCCCGCAATGGTTTATCA							
RIGHT PRIMER	694	22	62.63	50.00	4.00	2.00	
GTAAAGTTGGCAGTCGCAGACA							
PRODUCT SIZE: 680, PAIR ANY COMPL: 4.00, PAIR 3' COMPL: 1.00							
4 LEFT PRIMER	13	22	59.25	40.91	3.00	2.00	
GTTATCTCCCGCAATGGTTTAT							
RIGHT PRIMER	691	19	60.61	52.63	4.00	2.00	AAGTTGGCAGTCGCAGACA
PRODUCT SIZE: 679, PAIR ANY COMPL: 3.00, PAIR 3' COMPL: 1.00							

Statistics

	con sid ered	too many Ns	in tar get	in excl reg	bad GC%	no GC clamp	tm too low	tm too high	high any compl	high 3' compl	poly X	high end stab
ok												
Left	302	0	0	0	0	0	40	188	2	8	0	11
53												
Right	335	0	0	0	0	0	16	274	0	7	0	6
32												

damX down fw: TTATCTCCCGCAATGGTTTATC

damX down rev:TAAAGTTGGCAGTCGCAGACA