OGRE cycle 1

- 1. Add 100 ul of electroporation competent cells to ice cold electroporation cuvettes.
- 2. Add 5uM oligo to electroporation cuvette and was mixed gently with pipetting or flicking.
- 3. The sample was Incubated for 5 10min.
- 4. Prepare 1 sterile plastic tube for each sample with 1 ml recovery medium, put them on ice
- 5. Sample was added to electroporation's cuvette and electroporated with 2.2kV (for 0.2 cm cuvettes). Runtime should be 5-6ms. Optimal runtime is 5.5.
- 6. Immediately after the electroporation add 1 ml of recovery medium to the electroporation cuvette and pipette this to a sterile plastic tube. Repeat this 2 times. Now the total volume should be about 3 ml in each plastic tube.
- 7. Incubate the samples for 4h at 30 degC shaking at 220rpm.
- 8. Optional: measure OD_{600} of sample and use following equation to calculate the a suitable dilution.
 - a. $d = log_{10}(1.33 * X_{OD} * 10^5)$, where the right dilution will be 10^{d} .
 - b. This equation will over estimate the dilution, so dilution of 10^{d-1} should be used as well.
- 9. Plate the samples on appropriate antibiotics and in appropriate dilutions.
- 10. If doing more cycles save the rest of the sample and continue to "More cycles of OGRE".

More cycle of OGRE

- 11. Centrifuge samples at 5000g for 10 min. at 4 degC
- 12. Dispose the supernatant and resuspend in 1/20 (of original volume) cold electroporation medium. Repeat this 4 times. Keep everything cold.
- 13. Go to step 1.

References

This is a modified version of the protocol from Sun et. al, 2015 [1]. Modifications was inspired from Lu et al, 2012 [2] and Carr et al, 2012 [3].

¹ See http://2015.igem.org/Team:DTU-Denmark/Project

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