DNA laddering

I 、Analytical Gel

1) Composition

-10×TAE	60ml
-Biowest Agarose	0.9g
-SYBRSafe	0.6µL

2) Boil over med-high hit for 1.5 min until the powder completely soluted. Add SYBRSafe(6µL).

3) Pour 30ml solution in per small gel tray. Place 11- well comb. Wait for the gels to solidify (20min) .

 $1 \square$ Running the gel

- Put the gel with the gel tray in enough 10xTAE.

- Load 2µL Marker in one plate holder.

-Add $3\mu L$ FD Green buffer to the DNA. Load $3\mu L$ into each plate holder.

-Set the timer and voltage to 30min 125V.

2, Autoradiography and Annotation

II 、Extraction Gel

1, Agarose gel

1) Composition

-10×TAE 60ml

-Agarose 0.9g

-SYBRSafe 0.6µL

2) Boil over med-high hit for 1.5 min until the powder is completely soluted. Add SYBRSafe (6µL).

3) Pour 30ml solution in per small gel tray. Place 6-well comb. Wait for the gels to solidify (20min) .

2 Running the gel(Marker:6µL)

3 Cut out the appropriate bands. Place into Eppendorf tubes.

 $4\square$ Extraction

- Add 500µL BL into CA2.Spin 12000rpm for 1min.

Use equal volume Buffer PN to dissolve the gel. Incubate at 55°C 1300rpm until the it is completely dissolved.
Add dissolved gel into CA2 column in the collection tube. Wait for 2min at 25°C.Spin at 12000rpm for 1min.
Discard the liquid in the collection tube. Repeat..

- Add 600µL Buffer PW into the CA2 column. Spin at 12000 rpm for 1min. Discard the liquid in the collection tube.

- Spin the empty column in collection tube at 12000rpm for 1 min. Discard the collection tube.
- Put the CA2 column in an Eppendorf tube. Incubate at 55°C for 5min with the cap open.
- Add 50µL Buffer EB. The buffer should be pipetted onto centre of filter .Incubate at 55°C for 2min. Then spin at

12000rpm for 2min.

- Measure the concentration of DNA.