

## Growing Medium of *E.coli* :

### Overview :

We used LB broth (Luria broth, Luria-Bertani medium) for plasmid DNA production (cloning, transformation and miniprep). Otherwise, we used only M63 media for curdian production because it is a minimal medium, so we could to control all media parameters for production.

### Media :

#### - Luria-Bertani (LB) broth: 1L

Component	Volume & Mass	Procedure
Bactotryptone	10 g	1) Adjust pH to 7.5 with NaOH 2) Adjust volume to 1 L 3) Sterilize by autoclave
Yeast Extract	5 g	
NaCl	10 g	

- For LB plates : add 12 g/L of agar

#### - Preparation of 5X M63 Medium

Component	Volume & Mass	Procedure
(NH <sub>4</sub> ) <sub>2</sub> SO <sub>4</sub>	10 g	1) Add the following reagents to a 2-liter flask 2) Adjust volume to 1 L 3) Once the ingredients are added, heat with stirring until the components are completely dissolved.
KH <sub>2</sub> PO <sub>4</sub>	68 g	
FeSO <sub>4</sub> .7H <sub>2</sub> O	2,5 mg	
		4) Adjust pH to 7.0 with Acid 5) Sterilize by autoclave

#### - Preparation of 1X M63 Medium Working Solution

Aseptically dilute 200mL of 5X stock solution with 789 mL of sterile distilled water.

Aseptically add the following sterile solutions:

- 1 mL of 1 M MgSO<sub>4</sub>.7H<sub>2</sub>O (*directly in the 1X medium, not in the 5X*)
- 10 mL of 20% carbon source (*final concentration: 0,2 %*)
- 0.1 mL of 0.5% vitamin B1 (thiamine)  
« Vitamins should be added to a final concentration of 1µg/mL or 1mg/L»
- Antibiotic

#### Optional :

Add 5 mL of 20% Casamino Acids or L amino acids to 40 µg/mL or DL amino acids to 80 µg/mL

#### - Preparation of Stock Carbohydrate Solution (Glucose) :

Add 20 g of carbohydrate to distilled water and bring volume to 100mL.

Mix Thoroughly.

Filter sterilize.

#### - Preparation of Stock MgSO<sub>4</sub>-7H<sub>2</sub>O Solution :

Add 24,65 g of MgSO<sub>4</sub>-7H<sub>2</sub>O to distilled water and bring volume to 100mL.

Mix Thoroughly.

Filter sterilize.

**Antibiotic :**

Antibiotic	Stock concentration	Working concentration	Dissolve in
Ampicillin	50 mg.mL <sup>-1</sup>	50 µg.mL <sup>-1</sup>	H <sub>2</sub> O
Kanamycin	50 mg.mL <sup>-1</sup>	50 µg.mL <sup>-1</sup>	H <sub>2</sub> O
Chloramphenicol	34 mg.mL <sup>-1</sup>	10 µg.mL <sup>-1</sup>	95% Ethanol
Tetracycline	12,5 mg.mL <sup>-1</sup>	12,5 µg.mL <sup>-1</sup>	50% Ethanol

- Cool down the medium to 50°C before adding antibiotics.

**Curdlan Production with *E.coli* :**

1. Pre-warming medium at 37°C to decrease the time of lag-phase
2. Take colonies and inoculate 50 mL of complete M63 (1X) + Antibiotic at 37°C overnight
3. Grow cells until A600 : 0.7-0.9
4. Inoculate 30 mL inoculum for 120 mL of the M63 complete medium + antibiotic in a 500 mL Erlenmeyer flask (in order to have A600: 0.2)
5. Put the flask at 37°C, 180 rpm
6. Take out 10 mL of culture, centrifuge 5min at 14,000 rpm and 4°C, discard supernatant and store pellet at -20°C (this is the uninduced time point).
7. During stationary phase, re-incubate remaining cultures at 25°C shaking with 180rpm during 21h.