

The DNA extraction with alkaline extraction

1.5 ml tube

↓

+ 1.5 ml culture

↓

Centrifuge 13000 rpm 1min 4°C

↓

Collect the supernatant and cast aside

↓

+ Soln 1 200 μ l mix by voltex

↓

+ soln 2 200 μ l

↓

Gently mix by inverting the tube

↓

+ soln 3 200 μ l

↓

Gently mix

↓

On ice 5min

↓

Centrifuge 13000 rpm 10 min 4°C

↓

Collect the supernatant and add to the new tube

↓

+ phenol, chloroform, isoamylalcohol 500 μ l

↓

Mix by voltex

↓

Centrifuge 13000 rpm 5 min 4°C

↓

Take only the only water layer

↓

add to the new tube

↓

+100 % Etoh 400 μ l
 ↓
 Mix by vortex
 ↓
 Centrifuge 13000 rpm 20 min 4 °C
 ↓
 Throw away the supernatant to alcohol waste bin
 ↓
 +70 % Etoh 500 μ l
 ↓
 Gently mix
 ↓
 Throw away the supernatant to alcohol waste bin
 ↓
 65 °C to dry
 ↓
 +TE buffer 50 μ l
 ↓
 +RNase A (0.5 μ g/ μ l)
 ↓
 37 °C 20 min
 ↓
 Vortex and Tabletop small centrifuge
 ↓
 4 °C preserve Refrigerator

TE buffer 1ml (dH₂O 985 μ l, 1M Tris-HCl (pH8.0) 10 μ l, 200mM EDTA 5 μ l)
 Soln 1 1 ml (dH₂O 900 μ l, 1M Tris-HCl (pH8.0) 50 μ l, 200mM EDTA 50 μ l)
 Soln 2 1 ml (dH₂O 876 μ l, 5M NaOH 24 μ l, 10%SDS 100 μ l)
 Soln 3 1 ml (dH₂O 290 μ l, 5M potassium acetate , 600 μ l, acetic acid 110 μ l)
 RNase A (0.5 μ g/ μ l) 1 μ l

1M Tris-HCl 60 μ l: 3¥ \cong 0.03 \$
 200mM EDTA 55 μ l: 0.396¥ \cong 0.003 \$
 5M NaOH 24 μ l: 0.028¥ \cong 0.0002 \$
 10%SDS 100 μ l: 1.8¥ \cong 0.0149 \$

5M potassium acetate 600 μ l : 1.65 ¥ \doteq 0.0136 \$

acetic acid 110 μ l : 0.18 ¥ \doteq 0.0015 \$

RNase A (0.5 μ g/ μ l) 1 μ l : 130 ¥ \doteq 1.1 \$

Tube \times 3 : 6.5 ¥ \doteq 0.054 \$

TOTAL 144 ¥ \doteq 1.22 \$