12.8 Interlab measurement

WEDNESDAY, 8/12

Made the transformation for four constructs into TOP10: D1 A, D2, AHD6 and AHD7. Did also the same protocol for component cells without adding any plasmid in order to make +control for measurements.

Added 50 ul of ice cold compenents cells into pre-chilled 2ml tube and 2 ul of resuspended DNA to the same tube, and mixed with pipette gently. Liquid was transparent and there was some moisture on the tube walls. The cells were incubated on ice for 30 mins. The tubes were put in a water bath (42C) for 60s. After the heat shock the cells were on ice incubation for 5 minutes. Incubated the cells at 37C for 1 hr with shaking (230 RPM).

17.8.2015 Interlab Measurement

MONDAY, 8/17

Started o/n for each sample (D1, D2) and control (pSB1C3, one without any plasmid and J23151) of triplicate at 17.25 (16-18h, 300rpm, 37C) with 3ml of LB in culture tubes. Tubes taken out at 10.35. (Add more tube information) All the medias had the chloraphenicol concentration of 35 ug/ml expect for the cells which didn't contain any plasmids.

18.8.2015 Interlab Measurement

TUESDAY, 8/18

Measured yesterday's OD600 for samples and diluted until the OD600 was 0,5 (Within 5% accuraty) . The following results were obtained:

Table1

Sample	OD600 at the beginning	OD600 after the dilution
D1a	2,1295	0,4821
D1b	2,3121	0,4994
D1c	2,1149	0,5175
D2a	2,6414	0,4921
D2b	2,6627	0,4831
D2c	2,5673	0,5027
AHD6a	2,3522	0,4781
AHD6b	2,5068	0,4883
AHD6c	2,3597	0,5229
AHD7a	2,2740	0,5185
AHD7b	2,3596	0,5124
AHD7c	2,1406	0,5139
TOP10a	2,5202	0,4787
TOP10b	2,4730	0,4924
TOP10c	2,2122	0,4927

Samples were kept in +4C for 1hr and 200ul media were transfered into a 96-well plate for each case. The fluoresence emission was measuder with Excitation max of 470nm and Emission max of 511nm. The results can be seen in http://2015.igem.org/Team:Aalto-Helsinki/InterLab.

26.8.2015 Interlab Measurement study

WEDNESDAY, 8/26

Anna restricted D1 (5.8., 51,9ng/µl) and D2 (3.7., 169,0ng/µl) with EcoRI for gel analysis. Used 3.1. restriction buffer and followed the restriction protocol. Tuukka continued wtih inactivation.

Ran samples in the gel electrophoresis (110V, 30 min, 1,25% agarose gel). The results can be seen in fig 1.

Geldoc_2015-08-26_18hr_04mini_interlab.png

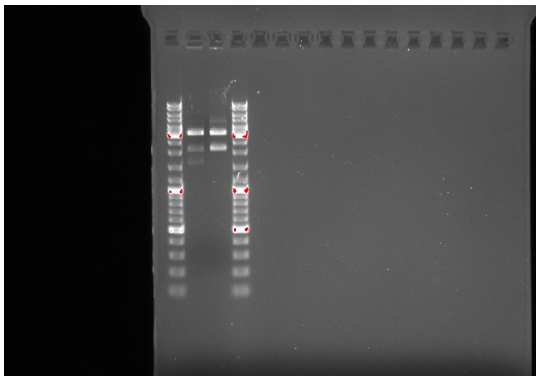

Figure 1. Restriction Mapping for D1.

The samples seems to be slight bigger (~3100 bp) than expected (2989 bp).