

## **Purification of 6xHis epitope tagged proteins by Ni-NTA-Agarose His**

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Low ( "Low" ) Imidazole Buffer 0.5L

100mM Imidazole 3.4g

5% glycerol 25ml 100% glycerol

50mM Tris-HCl (pH 7.9) 50ml 0.5M Tris-HCl (pH 7.9)

0.1% Tween-20 0.5M 100% Tween-20

500mM NaCl 50ml 5M NaCl

dH<sub>2</sub>O Fill to 0.5L (start w/ 350ml)

High ( "High" ) Imidazole Buffer 0.5L

500mM Imidazole 17.0g

5% glycerol 25ml 100% glycerol

50mM Tris-HCl (pH 7.9) 50ml 0.5M Tris-HCl (pH 7.9)

0.1% Tween-20 0.5M 100% Tween-20

500mM NaCl 50ml 5M NaCl

dH<sub>2</sub>O Fill to 0.5L (start w/ 350ml)

Sonication and Solubility Test:

Resuspend 1L worth of bacterial pellet in 30ml Low Buffer

Take 30  $\mu$ l pre-sonication sample

Sonicate 3 pulses at 80% power with 7th floor sonicator, 2 min on ice between pulses (NOTE: do not let sonicator tip touch side of tube to reduce frothing)

Take 30  $\mu$ l crude sonicated sample

Dispense into 1.5 ml eppendorf tubes, spin 14k rpm, 4°C, 30 min

Combine soluble fractions into 1 tube

Take 30  $\mu$ l soluble fraction as post-sonication soluble sample

Test solubility of target protein by SDS-PAGE/Coomassie Blue Stain

Add 10  $\mu$ l 4x Sample Buffer and 2  $\mu$ l  $\beta$ -Mercaptoethanol to each 30  $\mu$ l sample

Add an equal volume of 4x Sample Buffer to one insoluble pellet, add

$\beta$ -Mercaptoethanol to a final concentration of 5%.

Load 14  $\mu$ l (equals 10  $\mu$ l of fraction) on an appropriate concentration SDS-PAGE gel, electrophorese to separate bands well

Stain with Coomassie Blue for 30 min @ RT with rocking, destain with shreds of brown paper towel to aid in removal of dye from gel