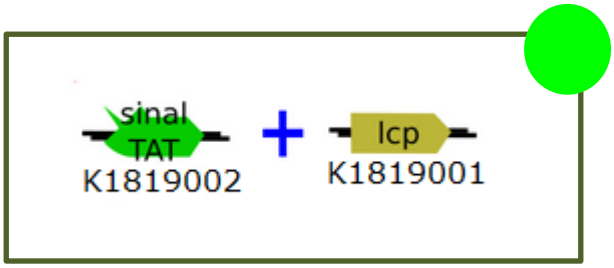
Assembly:

TAT_Lcp



1st Day:

EXSP Digestion (see **Enzymatic Digestion Protocol**)

	Part	Size	ηg/µl
1	K1819001	1138 bp	125
2	K1819002	147 bp	112

	Volume to 1,0 µg (µl)	Buffer 10x (µl)	Enzime 1	Volume (µl)	Enzime 2	Volume (µl)	H_2O to 20µl (µl)
1	8	2 (Tango)	X	1	SacI	1	8
2	7	2 (Tango)	X	1	SacI	1	9

Final Plasmid	Resistence
pSB1C3	chloramphenicol

Gel purification

- See PureLink® Quick Gel Extraction Kit Invitrogen™ manual
- Quantify digestion products

Parts	ηg/µl
K1819002	7.6
K1819001	2.9

Obs: 260/280 in a quality parameter that tells you if your sample is contaminated with proteins. The greater it is compared to 1 the less contaminants you have.

Ligation (see **Ligation Protocol**)

Part containing the plasmid	K1819001	6.5 µl
Insert	K1819002	4.2 µl
10x T4 DNA Buffer	2 µl	
T4 DNA ligase 1u	0.5 µl	
H2O to 20µl	6.8 µl	

Obs: To determinate the amount of DNA necessary we used the following equation

$$\textit{Insert ng} = \textit{plasmid ng} \times \textit{insert bp plasmid bp} \times \textit{insert: plasmid ratio}$$

- Incubate overnight at 37°C.
- Prepare and sterilize in the autoclave tubes with 6 ml of liquid LB medium.
- Prepare glycerol 40%

2nd Day:

Transformation (see **Transformation Protocol in Escherichia coli DH5-α**)

- Organism: E. coli DH5-α
- Selection: Chloramphenicol

4th Day:

Confirmation with NotI