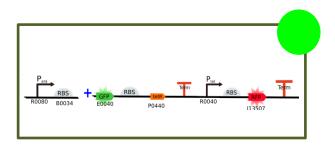
Assembly:

Pa_I13507



1st Day:

EXSP Digestion (see Enzymatic Digestion Protocol)

| | Part | Size | ηg/µl |
|---|------------|---------|-------|
| 1 | Pa_RBS | 161 bp | 94.8 |
| 2 | GFP_I13507 | 2585 bp | 273.5 |

| | Volume to 1,0 μg (μl) | Buffer 10x (µl) | BSA | Enzime 1 | Volume (µl) | Enzime 2 | Volume (µl) | H2O to 20µl (µl) |
|---|--------------------------|--------------------|-----|----------|----------------|----------|----------------|---------------------|
| 1 | 10 | 2 (M) | - | SpeI | 1 | PstI | 1 | 6 |
| 2 | 4 | 2 (M) | - | XbaI | 1 | PstI | 1 | 12 |

| Final Plasmid | Resistence |
|---------------|------------|
| pSB1A2 | ampicillin |

Gel purification

- See PureLink® Quick Plasmide Miniprep Invitrogen™ manual
- Quantify digestion products

| Parts | ηg/μl |
|------------|-------|
| Pa_RBS | 14 |
| GFP_I13507 | 12.3 |

Obs: 260/280 in a quality parameter that tells you if your sample is contaminated with proteins. The greater it is compared to 1 the less contaminants you have.

Ligation (see Ligation Protocol)

| Part containing the plasmid | Pa_RBS | 4 µl |
|-----------------------------|------------|---------|
| Insert | GFP_I13507 | 10.5 µl |
| 10x T4 DNA Buffer | 4 µl | |
| T4 DNA ligase 1u | 0.5 µl | |
| H2O to 20µl | 1 | |

Obs: To determinate the amount of DNA necessary we used the following equation

Insert $ng = plasmid ng \times insert bp plasmid bp \times insert: plasmid ratio$

- Incubate overnight at 37°C.
- Prepare and sterilize in the autoclave tubes with 6 ml of liquid LB medium.
- Prepare glycerol 40%

2nd Day:

Transformation (see Transformation Protocol in Escherichia coli DH5-α)

- Organism: E. coli DH5-α
- Selection: Ampcillin

4th Day:

Confirmation with NotI