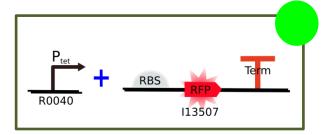
# Assembly:

Pt\_I13507



## 1st Day:

## EXSP Digestion (see Enzymatic Digestion Protocol)

	Part	Size	ηg/μl
1	R0040	164 bp	78.3
2	I13507	861 bp	214

	Volume to 1,0 μg (μl)	Buffer 10x (µl)	BSA	Enzime 1	Volume (µl)	Enzime 2	Volume (µl)	H2O to 20μl (μl)
1	13.5	2 (M)	-	S	1	P	1	2.5
2	4.5	2 (M)	-	X	1	P	1	11.5

Final Plasmid	Resistence
pSB1C3	chloramphenicol

## Gel purification

- See PureLink® Quick Plasmide Miniprep Invitrogen™ manual
- Quantify digestion products

Parts	ղց/μl
R0040	8,6
I13507	9.8

**Obs:** 260/280 in a quality parameter that tells you if your sample is contaminated with proteins. The greater it is compared to 1 the less contaminants you have.

## 3<sup>rd</sup> Day:

#### Ligation (see Ligation Protocol)

Part containing the plasmid	R0040	6 µl
Insert	I13507	6 µl
10x T4 DNA Buffer	4 μΙ	
T4 DNA ligase 1u	0.5 μ1	
H2O to 20µl 3.5		.5

**Obs:** To determinate the amount of DNA necessary we used the following equation

#### Insert $ng = plasmid ng \times insert bp plasmid bp \times insert: plasmid ratio$

- Incubate overnight at 37°C.
- Prepare and sterilize in the autoclave tubes with 6 ml of liquid LB medium.
- Prepare glycerol 40%

#### 4<sup>th</sup> Day:

Transformation (see Transformation Protocol in Escherichia coli DH5-α)

- Organism: E. coli DH5-α
- Selection: Ampcillin

#### 5<sup>th</sup> Day:

Confirmation with NotI