

Miniprep

2 mL of LB medium (supplemented with the appropriate antibiotic) with bacterial culture were inoculated and were left incubating overnight at 37 °C with agitation at 225 rpm. After the incubation period was finished, 1.5 mL of culture were deposited into a microcentrifuge compatible 2 ml tube and centrifugated for 30 seconds at 4 °C. Once the centrifugation was finished, the supernatant was discarded and resuspended in 100 µL of lysis solution 1, then it was vortexed for 5 min. Afterwards 200 µL of lysis solution 2 was added into the tube and mixed through inversion, then it was left to incubate for 15 min on ice. 150 µL of lysis solution 3 was then added into the tube and mixed softly through inversion to be left for a 15 min incubation on ice once again. Once the incubation period was completed, the tube was centrifugated for 5 min at maximum velocity. The supernatant was recovered in a new tube and an equal volume of phenol:chloroform:isoamyl alcohol 25:24:1 was added. The contents were then mixed by vortex and then centrifugated at max velocity in the microcentrifuge. The aqueous phase was transferred into a new tube, and mixed with 2 volumes of 96% ethanol at room temperature and vortexed once more, then it was left incubating for 2 minutes at room temperature. The contents were centrifuged once more at maximum velocity for 5 min at 4 °C and the supernatant was discarded. The tube was then left to dry open, inverted and over a towel until the remaining ethanol evaporated. 1 mL of 70% ethanol was then added, mixed by inversion and then the contents were centrifugated for 5 min in maximum velocity once more. The contents were left to dry just as before open, inverted and over a towel until the ethanol had evaporated. The extracted nucleic acids were then resuspended on 50 µL of TE (pH 8.0) and gently vortexed for a few seconds. This solution was then stored at -20 °C until it was used.